ORIGINAL ARTICLE

Friedelin inhibits the growth and metastasis of human leukemia cells via modulation of MEK/ERK and PI3K/AKT signalling pathways

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Summary

Purpose: Leukemia constitutes just 3% of all malignancies but because of its high incidence and mortality in children and persons below 40 years of age, it is considered one of the devastating malignant conditions. This study was undertaken to evaluate the anticancer effects of Friedelin triterpenoid against the human AML-196leukemia cells.

Methods: CCK-8 assay was used to determine cell viability. DAPI staining was used for the assessment of apoptosis. Annexin V/propidium iodide (PI) analysis was employed for the detection of percentage of apoptosis. Transwell assays were used for cell migration and invasion. Western blotting was used for the determination of protein expression.

Results: The results showed Friedelin inhibited the proliferation of the human AML-196 cells with little effects on the liferation

normal cells. Investigation of the underlying mechanisms showed that Friedelin induced apoptosis in AML-196 cells. Friedelin-induced apoptosis was linked with upregulation of cleaved caspase-3, 8 and 9, as well as cleaved PARP. The Bax protein levels were increased and of Bcl-2 were decreased. Transwell assays showed that Friedelin suppressed the migration and invasion of the AML-196leukemia cells. Additionally, Friedelin also blocked the MEK/ERK and PI3K/AT signalling cascades dose-dependently.

Conclusion: Taken together, Friedelin may prove beneficial in the treatment of leukemia.

Key words: leukemia, apoptosis, cell cycle, Friedelin, pro-

Introduction

cope up with the changing environmental conditions, they synthesize a huge number of natural *folia* to name a few [4,5]. Studies have shown that compounds. Triterpenoids have shown tremendous health promoting benefits [1]. Studies have shown that many of the triterpenoids suppress the growth of human cancers [2]. The triterpenoids derived from plants have been used in the treatment of different human disorders in Asian countries [3].

Plants are sessile organisms and in order to Friedelin has been isolated from several plant species such as Azimatetracantha and Maytenusilici-Friedelin suppresses the growth of glioblastoma cells [6]. Although leukemia constitutes just 3% of all malignancies but because of its high incidence and mortality rate and high incidence in children and persons below 40 years of age, it is considered one of the devastating conditions [7,8]. Only few

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efficient chemotherapeutic agents are available for leukemia and they too have severe side effects [9]. With this background, the identification of novel therapeutics may enable the proper management of leukemia. Given mortality, there is high urgency to identify compounds that are safer and effective in the treatment of leukemia.

The present study reports that Friedelin significantly suppressed the growth of the human leukemia via induction of apoptosis. This molecule also suppressed the migration and invasion of AML-196 leukemia cells via blocking of the MEK/ERK and PI3K/AKT signalling pathway. Taken together, Friedelin may prove beneficial in the treatment of leukemia.

Methods

Growth and proliferation assay

Cell counting kit-8 (CCK-8, MedChemExpress) was used for estimation of the proliferative rates of AML-196 leukemia cells treated with Friedelin (Sigma-Aldrich, St. Louis, Missouri, United States) which were compared with those of HS-5 normal bone marrow cell lines. In brief, the cells were placed in 96-well plates at 1×10^6 cells/well and cultured for 24h with 0, 2.5, 5, 10, 20, 40, 80 and 160µM Friedelin, after which CCK-8 was employed to estimate the proliferation rates by the addition of 10µl of CCK-8 solution to each well, at the indicated time intervals. Following 2-h incubation at 37°C, absorbance at 450 nm was read for each sample with of microplate reader.

Apoptosis assay

DAPI staining was used to examine thse effect of Friedelin on the viability of leukemia cells and induction of cell apoptosis. The cells were placed in 12-well plates at a density of 0.6×10^6 cells/well. Friedelin at the concentrations of 0, 5, 10 and 20 µM was added to each well and cells were incubated at 37°C for 24 h. Afterwards, the cells were harvested and washed twice with phosphate buffered saline (PBS), followed by fixing with

4% paraformaldehyde. The DAPI solution was then used to stain the cells. Afterwards, the cells were examined for fluorescence measurements using fluorescent microscope. Annexin V/PI staining was used as described previously [3] to determine the percentage of apoptotic cells.

Migration and invasion assay

Transwell chamber with or without Matrigel coating was used to assess the migration and invasion of AML-196 leukemia cells. Briefly, 100µl cell culture containing 6000 cells was added to the upper chamber of the transwell and the lower chamber was added with 750µl of DMEM supplemented with 10% fetal bovine serum (FBS). After 48-h incubation at $37^{\circ}C/5\%$ CO₂, cells from the surface of membrane's upper side were removed carefully with cotton swabs while those sticked to lower side of membrane were fixed with 70% ethyl alcohol and stained with 0.1% crystal violet. Light microscope (×100) was used for visualization of cells and photographs were taken. At least seven random fields were used for counting of migratory or invasive cells.

Western blotting

Using RIPA lysis buffer (Thermo Fisher Scientific, Waltham, Massachusetts, United States), total proteins were isolated from untreated leukemia cells and cells treated with 10, 20 and 40µM Friedelin for 24 h. Bradford method was used to quantify the protein concentrations. About 45 µg of total proteins from each sample were separated electrophoretically on 10% SDS-PAGE. The gel was blotted to nitrocellulose membrane which gave the exposure of primary protein antibodies followed by exposure of secondary antibodies. Enhanced chemiluminescence (ECL) reagent was used for the detection of bands corresponding to proteins of interest. The protein expression was normalized with GADPH protein.

Statistics

The experiments were performed in triplicate and expressed as mean \pm SD. One-way ANOVA followed by Turkey's test were performed using SPSS software. P value < 0.05 was considered as showing statistically significant difference.



Figure 1. A: Chemical structure of Friedelin. **B:** CCK-assay showing the effect of Friedelin on the viability of Leukemia AML-196 and Hs-5 cells. The experiments were performed in triplicate and expressed as mean ± SD (*p<0.05).

Results

Friedelin suppresses the growth of leukemia cells

Friedelin (Figure 1A) exerted inhibitory effects on the proliferation of leukemia cells as ascertained by CCK-8 assay. The normal Hs-5 bone marrow cells and AML-196 leukemia cells were treated with 0 to 160 μ M of Friedelin for 24 h and it was found that the viability of AML-196 cells decreased proportionally with the increasing doses





Figure 2. DAPI staining showing the effect of Friedelin on the nuclear morphology of the AML-196 cells at varied doses. The experiments were performed in triplicate.



Annexin V-FITC

Figure 3. Annexin V/PI staining showing the effect of Friedelin on the percentage of apoptosis of the AML-196 cells at varied doses. The experiments were performed in triplicate.

of Friedelin with an IC₅₀ of 10 μ M (Figure 1B). Surprisingly, the effects of the molecule on the normal NCI-H526 cells were less severe as evidenced from the IC₅₀ of around 80 μ M.

Friedelin promotes apoptosis in AML-196 leukemia cells

The DAPI staining revealed that the suppressive effects of Friedelin on the proliferation of AML-196 human leukemia cells were mainly because of the initation of apoptosis as evidenced from the increase in the nuclear fragmentation (Figure 2). The extent of apoptosis induced by Friedelin was determined by annexin V/PI staining. The percentage of apoptosis increased from 3.8% to 27.8% with 20 µM Friedelin compared to control cells (Figure 3).

Friedelin alters apoptosis-related protein expression

Friedelin-induced apoptosis was also confirmed by examining the effects of the expression of the apoptosis-related proteins. The results showed a concentration-dependent increase in the proteins levels of cleaved caspsase-3, 8 and 9 as well as of the cleaved PARP in Friedelin-treated AML-196 cells. Furthermore, the expression of Bax increased while of Bcl-2 showed a dose-dependent decrease (Figure 4).

Friedelin suppresses migration and invasion of AML-196 cells

The determination of the effects of Friedelin on the migration and invasion of the human AML-196 leukemia cells was done by transwell assays.



Figure 4. Western blot analysis showing the effects of Friedelin on the expression of apoptosis-related proteins. The experiments were performed in triplicate.

The results revealed that the molecule suppressed the migration of the AML-196 cells concentrationdependently (Figure 5). The effects of this molecule were also determined on AML-196 cell invasion and the results were similar to cell migration (Figure 6).

Friedelin blocks MEK/ERK and PI3K/AKT signalling

The impact of Friedelin were also evaluated on the MEK/ERK signalling pathway. It was found



Figure 5. Effect of Friedelin on the migration of the AML-196 leukemia cells. The experiments were performed in triplicate.



Figure 6. Effect of Friedelin on the invasion of the AML-196 leukemia cells. The experiments were performed in triplicate.

that the protein levels of p-MEK and p-ERK declined significantly and concentration-dependently (Figure 7). Additionally, it was revealed that the molecule caused concentration-dependent inhibition of phosphorylation of PI3K and AKT (Figure 8). Nonetheless, there was no apparent effect on the total PI3K and AKT.

Discussion

During the course of evolution plants have learned to defend themselves by synthesising a wide array of molecules which also include trit-



Figure 7. Western blots showing the effect of Friedelin on the MEK/ERK signalling pathway. The experiments were performed in triplicate.



Figure 8. Western blots showing the effect of Friedelin on the PI3K/AKT signalling pathway. The experiments were performed in triplicate.

erpenoids. Triterpenoids constitute a large and diverse group of plant-derived compounds with enormous pharmacological potential [10] and they ubiquitously present across the plant kingdom. These plant-derived metabolites have a wide array of biological roles in plants which include defense against the biotic and abiotic stresses. Accordingly, active research is going on to examine the anticancer effects of triterpenoids against different human cancers. This study was undertaken to evaluate the anticancer effects of Friedelin, a naturally occurring triterpenoid of plant origin against the human leukemia cells. The cell proliferation assay showed significant inhibition of the leukemia cells growth upon Friedelin treatment. Previous studies have shown that triterpenoids have the potential to trigger apoptosis in the leukemia and neuroblastoma cells [11].More precisely, Friedelin has been shown to suppress the growth of breast cancer cells via induction of apoptosis [12]. The expression of cleaved caspase-3, 8 and 9 as well as that of cleaved PARP was remarkably increased. Additionally, the Bax/ Bcl-2 ratio was also increased which is an important indicator of apoptosis [13]. Apoptosis plays key role in eliminating the defective cells and thus drugs that promote apoptosis are currently being studied extensively [14]. This study also examined the effects of Friedelin on migration and invasion of the AML-196 cells and found that this molecule suppressed their migration and invasion. These findings are in agreement with a previous study wherein a triterpenoid Ursolic acid has been found to suppress the migration and invasion of cancer

cells [15]. MEK/ERK and PI3K/AKT signalling pathways are among the critical signalling cascades that are dysregulated in cancer cells [16]. Herein we observed that Friedelin blocks both the MEK/ERK and PI3K/AKT signalling pathway. Hence, more studies are required to establish Friedelin as a lead molecule for the development of new therapeutic agents for leukemia.

Conclusion

Taken together, the results of the present study showed that Friedelin inhibited the growth of the human leukemia cells via induction of apoptosis and G2/M cell cycle arrest. Friedelin also suppresses the migration and invasion of the leukemia cells and may therefore prove beneficial in the treatment of leukemia.

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Conflict of interests

The authors declare no conflict of interests.

References

- 1. Huang CC, Hsu MC, Huang WC, Yang HR, Hou CC. Triterpenoid-rich extract from Antrodiacamphorata improves physical fatigue and exercise performance in mice. Evidence-Based Complement Altern.Med 2012;2012.
- Gao X, Deeb D, Jiang H, Liu Y, Dulchavsky SA, Gautam SC. Synthetic triterpenoids inhibit growth and induce apoptosis in human leukemia and neuroblastoma cells through inhibition of prosurvival Akt, NF-κB and Notch1 signaling. J Neuro-oncol 2007;84:147-57.
- Weng JR, Bai LY, Chiu CF, Hu JL, Chiu SJ, Wu CY. Cucurbitane triterpenoid from Momordica charantia induces apoptosis and autophagy in breast cancer cells, in part, through peroxisome proliferator-activated receptor γ activation. Evidence-Based Complem Altern Med 2013;2013.
- 4. Antonisamy P, Duraipandiyan V, Ignacimuthu S. Anti-

inflammatory, analgesic and antipyretic effects of friedelin isolated from Azimatetracantha Lam. in mouse and rat models. J Pharm Pharmacol 2011;63:1070-7.

- Queiroga CL, Silva GF, Dias PC, Possenti A, de Carvalho JE. Evaluation of the antiulcerogenic activity of friedelan-3β-ol and friedelin isolated from Maytenusilicifolia (Celastraceae). J Ethnopharmacol 2000;72:465-8.
- 6. Akihisa T, Tokuda H, Ichiishi E et al. Anti-tumor promoting effects of Friedelin-type triterpenoids and cytotoxic activity of karounidiol against human cancer cell lines. Cancer Lett 2001;173:9-14.
- Deschler B, Lübbert M. Acute myeloid leukemia: epidemiology and etiology. Cancer 2006;107:2099-107.
- 8. Fraumeni JF Jr. Clinical epidemiology of leukemia. In: Semin Hematol 1969;6:250-60.

- 9. Zhang L, Freeman LE, Nakamura J et al. Formaldehyde and leukemia: epidemiology, potential mechanisms, and implications for risk assessment. Environ Molec Mutagenesis 2010;51:181-91.
- 10. Liby KT, Yore MM, Sporn MB. Triterpenoids and rexinoids as multifunctional agents for the prevention and treatment of cancer. Nat Rev Cancer 2007;7s:357.
- 11. Yadav VR, Prasad S, Sung B, Kannappan R, Aggarwal BB. Targeting inflammatory pathways by triterpenoids for prevention and treatment of cancer. Toxins 2010;2:2428-66.
- Subash-Babu P, Li DK, Alshatwi AA. In vitro cytotoxic potential of friedelin in human MCF-7 breast cancer cell: Regulate early expression of Cdkn2a and pRb1, neutralize mdm2-p53 amalgamation and functional stabilization of p53. Experim Toxicol Pathol 2017;69:630-36.

- 13. Lowe SW, Lin AW. Apoptosis in cancer. Carcinogenesis 2000;21:485-95.
- 14. Fisher DE. Apoptosis in cancer therapy: crossing the threshold. Cell 1994;78:539-42.
- 15. Yeh CT, Wu CH, Yen GC. Ursolic acid, a naturally occurring triterpenoid, suppresses migration and invasion of human breast cancer cells by modulating c-Jun Nterminal kinase, Akt and mammalian target of rapamycin signaling. Mol Nutrit Food Res 2010;54:1285-95.
- 16. Riquelme E, Behrens C, Lin HY et al. Modulation of EZH2 expression by MEK-ERK or PI3K-AKT signaling in lung cancer is dictated by different KRAS oncogene mutations. Cancer Res 2016;76:675-85.
- 17. Asati V, Mahapatra DK, Bharti SK. PI3K/Akt/mTOR and Ras/Raf/MEK/ERK signaling pathways inhibitors as anticancer agents: Structural and pharmacological perspectives. Eur J Med Chem 2016;109:314-41.