

REVIEW ARTICLE

Targeted treatment for metastatic renal cell carcinoma and immune regulation

K.A. Laschos¹, K.T. Papazisis^{1,2}, L.F. Kontovinis^{1,2}, C. Kalaitzis³, S. Gianakopoulos³,
A.H. Kortsaris⁴, S. Touloupidis³

¹Applied Molecular Oncology Laboratory and ²3rd Department of Medical Oncology, Theagenion Cancer Hospital, Thessaloniki; ³Department of Urology and Andrology and ⁴Laboratory of Biochemistry, Department of Medicine, University Hospital of Alexandroupolis, Democritus University of Trace, Alexandroupolis, Greece

Summary

New targeted agents have become the mainstream of treatment in metastatic renal cell carcinoma (mRCC) and substituted the previous cytokine-based therapies. Vascular endothelial growth factor (VEGF) pathway is the principle target for drugs like sunitinib, sorafenib and bevacizumab. As VEGF is regulating dendritic cell (DC) function, inhibition of VEGF results in activation of DCs and a shift towards cellular (type I) immunity, which is believed to favor cancer rejection. Recent studies have established the immune-stimulating effects of sunitinib that may as well be a marker for effectiveness. On the other hand, sorafenib not only inhibits VEGF receptor (VEGFR) but is also a B-Raf inhibitor (a component of the ras – MAPK pathway) and this leads to downregulation of immune responses. Sorafenib has not yet shown benefit in first-

line treatment of mRCC when compared to interferon (IFN)- α and sorafenib-mediated immunosuppression may partially account for that. Mammalian target of rapamycin (mTOR), the target of temsirolimus, is an element of the DC activation pathway. There are no data for in vivo effects of temsirolimus in the immune system. The addition of IFN- α to temsirolimus resulted in inferior outcomes than temsirolimus alone. IFN- α has however still a place in mRCC treatment, as bevacizumab has been approved in combination with IFN- α . New clinical trials address the effects of the combination of cytokines with targeted agents. The immune-modulating effects of targeted treatments may be important in pharmacodynamic outcomes, effectiveness or the development of adverse events.

Key words: immune regulation, renal cancer, sorafenib, sunitinib, temsirolimus

Introduction

RCC affects more than 50,000 patients in the United States each year, accounting for more than 13,000 deaths annually. These tumors account for approximately 3% of adult malignancies and occur in a male-female ratio of 1.6:1 [1]. The cell of origin, morphology and growth pattern characterizes RCC histology. Histologically, 4 major RCC subtypes have been identified: clear cell (60-80%), papillary (10-15%), chromophobe (5-10%) and collecting duct carcinoma (< 1%). Clear-cell histology is associated with a better outcome than papillary or chromophobe histology in the metastatic setting, but the opposite is true for localized disease [1,2]. Rare histologies like clear cell carcinoma with rhabdoid features have an even worse outcome [4]. Localized disease is curable with surgery but a third of patients present with incurable

metastatic disease. The aim of management (when metastatic disease is present) is palliation, although with the development of the novel targeted agents prolongation of life appears to be a real possibility. The median survival for patients with mRCC before the era of novel targeted treatments was 10-12 months [5]. After the introduction of targeted treatment (sunitinib), reports raise the median survival up to 2 years in patients that have previously failed cytokine therapy [6].

RCC, angiogenesis and immune surveillance

Angiogenesis is important for tumor growth and development as well as for metastatic spreading. RCC is often associated with deregulation of angiogenesis through alterations (mutation or gene methylation) of

the von Hippel-Lindau (VHL) tumor-suppressor gene [7], that regulates the expression of the hypoxia-inducible factor alpha (HIF- α). HIF- α is a transcription factor that controls the expression of several pro-angiogenic genes like VEGF and platelet-derived growth factor (PDGF) [8]. The VHL-HIF-VEGF axis is therefore a target for therapeutic interventions in mRCC [9].

DCs are the most potent antigen-presenting cells (APCs) [10] and play a central role in the host's anti-tumor immunity. Many investigators have described the defective function of DCs in tumor-bearing hosts [11,12]. One of the possible reasons for this DC defectiveness is the secretion of VEGF by tumor cells. VEGF is regulating immune responses by inhibiting DCs differentiation, maturation and function [13,14]. VEGF inhibits TLR4-mediated but not pro-inflammatory cytokine-mediated DC maturation [15]. This effect is differentially mediated by VEGF-receptor subtypes (VEGFR1 and VEGFR2) [16] and can be partially reversed by VEGF-trap [17]. Additionally, VEGF inhibits T-cell development and activation, and this may further contribute to tumor-induced immune suppression [18]. Inhibiting VEGF by bevacizumab or VEGFR by the protein tyrosine kinase inhibitors sunitinib and sorafenib may therefore reverse the DCs dysfunction and T-cell activation in patients with mRCC and/or synergize with immunotherapeutic approaches.

We will therefore discuss the rationale of such an approach and the emerging clinical and experimental data.

Treatment for metastatic RCC. From cytokines to targeted agents

Several lines of evidence suggest that RCC is an immune-sensitive cancer. For years, treatment with interleukin-2 (IL-2) or IFN- α was the only treatment available for patients with mRCC [19]. A minority of mRCC patients responds to IL-2 or IFN- α treatment with durable remissions that may even lead to cure in some of them, though this is at the expense of severe adverse reactions [20]. However, these agents may provide only modest increases in median survival [21]. IL-2 and IFN- α have now been largely replaced in the treatment of mRCC by novel agents targeting specific components of the pathways involved in tumor growth and angiogenesis [22]. VEGFR inhibition with the multitargeted receptor tyrosine kinase inhibitors sunitinib [23-25] (Sutent[®], Pfizer Inc.) and sorafenib [26] (Nexavar[®], Bayer HealthCare/Onyx Pharmaceutical) has proven to be an effective strategy for the treatment of mRCC and both agents are now in clinical practice and being tested in the adjuvant setting. Furthermore, the VEGF ligand-binding monoclonal antibody bevacizumab [27] (Avastin[®], Genetech, Inc.) and the mTOR kinase inhibitor temsirolimus (Torice[®], Wyeth Pharmaceuticals) have demonstrated clinical activity in patients with mRCC as well [28]. Recently, everolimus (Afinitor, RAD001, Novartis), another inhibitor of mTOR, was approved for the treatment of mRCC in patients progressing after therapy with sorafenib, sunitinib or both [29] (Table 1).

Table 1. Clinical data regarding 5 principal targeted agents used in everyday practice

<i>Treatment</i>	<i>ORR %</i>	<i>Any reduction in tumor burden, %</i>	<i>PFS</i>	<i>OS</i>
Immunotherapy				
High dose IL-2	20-23 [51-55]	NA	3.1 [54]	19 [53]
Low dose IL-2 or IFN- α	10-15 [56-59]	NA	4.7 [59]	12-14 [57]
Small molecular weight inhibitors of VEGFR				
Sunitinib	30-45 [23,24,60]	70-75	11 mos treatment-naïve [24] 8.4 mos cytokine-refractory [23,60]	26.4 mos [62] 21.8 mos [61]
Sorafenib	2-10 [26,61]	70-75	5.7 mos treatment-naïve [62] 5.5 mos treatment-refractory [26]	17.8 mos [63]
Bevacizumab (BEV) monotherapy	10-13 [64,65]	70-75	8.5 mos [65]	NA
BEV+IFN- α	26-31		10.2 mos treatment-naïve [27] 4.8 mos cytokine-refractory [64]	
mTOR inhibitors				
Temsirolimus	7-9 [28,66]	NA	3.7 mos treatment-naïve [28] 5.8 mos pretreated [66]	10.9 mos [28]
Everolimus	1 [29]	60 [29]	4.0 mos [29]	NA

ORR: objective response rate, PFS: progression-free survival, OS: overall survival, NA: not available, VEGFR: vascular endothelial growth factor receptor, IFN- α : interferon alpha, IL-2: interleukin 2, mTOR: mammalian target of rapamycin, mos: months

Cytokine treatment in the era of targeted therapies (Figure 1)

There is preclinical evidence suggesting that combining anti-VEGF therapy and immunotherapy leads to a greater antitumor activity than either agent alone in tumor-bearing mice [30]. IFN- α activates DCs [31] and it may synergize with bevacizumab in more effective stimulation of the immune responses. Interestingly, treatment with bevacizumab was approved in combination with IFN- α , as was the original design of the AVOREN study [27]. However, the AVOREN study had no arm without IFN- α and the benefit of bevacizumab was evident even when combined with very low IFN- α doses [32].

In the ARCC trial (the temsirolimus registration study) the combination of IFN- α with temsirolimus did not improve responses and resulted in inferior overall survival compared to the temsirolimus monotherapy arm [28]. We have to notice though that temsirolimus is

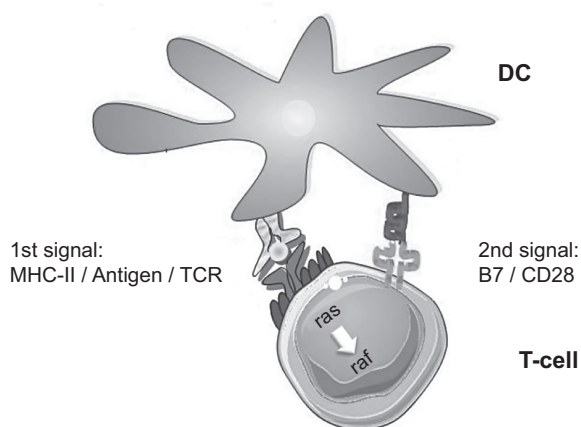


Figure 1. Schematic representation of the immune recognition by a T-cell. Dendritic cells (DCs) present antigen to T-cell receptor (TCR) via a major histocompatibility antigen class II (MHC-II). Downstream events include the activation of RAS – RAF – MAPK pathway, a target of sorafenib. On the other hand, effective antigen presentation requires at least a “second” signal that is usually given via a B7- CD28 interaction. Maturation of DCs and B7 expression is inhibited by VEGF, and sunitinib/sorafenib increase the expression of the costimulatory molecules in the surface of DCs. A third signal that polarizes T-cells towards a TH1 or a TH2 immune response may be regulated by the mTOR pathway.

not directly targeting VEGF as the other registered targeted agents. There are data showing that mTOR is crucial for the IFN responses of plasmacytoid DCs [33], the IL-18 production of DCs [34] and rapamycin can induce DCs apoptosis [35]. On the other hand, mTOR is regulating the TLR4-mediated IL-12 secretion by DCs and rapamycin increases IL-12 production (and cell-mediated immune responses) under these conditions [36]. The exact role of mTOR in DC physiology is still controversial but it seems that it is a component of the pathway that regulates the production of proinflammatory cytokines and shifts the maturation of DCs towards a more Th1-inducing type (high IL-12 and low IL-10 production) [37]. Inhibition of mTOR with temsirolimus and induction of inflammatory immune responses with IFN- α could stimulate Th2 humoral inflammatory responses, counting for the higher incidence of adverse reactions seen in the combination arm of the ARCC trial. Patients in the combination arm experienced more grade 3-4 adverse events and consequently had more delays and reductions in treatment.

The question as to what extent cytokine treatment may add effectiveness to targeted treatments remains still unanswered. However, it is intriguing to think that even though targeted therapies perform generally better than IFN- α , they do not lead to cure or the long-lasting remissions that a small minority of patients was experiencing with the cytokine treatment. Therefore, it would be important if it could be possible to identify those patients that may gain a large benefit from cytokine therapy and treat them with IFN- α or IL-2. Alternatively, new ways of combining targeted treatment with cytokines should be explored or attain deeper knowledge of the immune-mediating actions of targeted therapies. Such an approach may lead to better combinations or optimal sequencing and use of anti-mRCC treatments.

Do targeted treatments affect immune surveillance? (Table 2)

In concert with the *in vitro* data, sunitinib augments the *in vivo* immune responses. This is supported by nu-

Table 2. Targeted agents used in the treatment of metastatic renal cell carcinoma and their effects in immune regulation

Targeted agent	Indications	Immune effects
Sunitinib	1st line mRCC	Increases DCs numbers, maturation and function
Sorafenib	2nd line mRCC	Decreases DCs maturation and function, and T-cell function
Temsirolimus	1st line mRCC (after TKIs)	Probably immune suppression
Bevacizumab	1st line mRCC	In combination with IFN- α , probably IFN effects predominate (immune-stimulation)

mRCC: metastatic renal cell carcinoma, TKIs: tyrosine kinase inhibitors, DCs: dendritic cells, IFN- α : interferon alpha

Table 3. Markers expressed by dendritic cells and effect of targeted agents

Marker	Function	Effects of targeted agents
CD1a	Myeloid DCs	Increased by sunitinib in responders only, decreased by sorafenib
CD83	Maturation	Downregulated by sorafenib. Sunitinib increases CD83+ DCs in blood
CD80/CD86	Co-stimulation	Decreased by VEGF, reversed by bevacizumab and sorafenib (<i>in vitro</i> data). Lipopolysaccharide inductions decreased by sorafenib

DCs: dendritic cells, VEGF: vascular endothelial growth factor

merous recent studies that have addressed several aspects of the immune system in patients with mRCC treated with sunitinib. It has been shown that DC numbers increase to match normal controls after 4 weeks of sunitinib treatment and this is correlated with clinical responses [38]. T-cell activity increases and regulatory T-cells (T-regs) decrease [39]. Patients receiving sunitinib had a significant shift towards Th1 cytokine responses (increasing the intracellular IFN- γ /IL-4 ratio) when peripheral blood mononuclear cells (PBMCs) were stimulated *in vitro* with anti-CD3/anti-CD28 antibodies [40]. In the same cohort of patients, T-regs (CD4⁺CD25^{hi}) decreased after the administration of sunitinib. The immunostimulatory effects of sunitinib are probably due to the VEGFR inhibition; when the anti-VEGF monoclonal antibody bevacizumab is administered to patients with metastatic lung, breast or colorectal cancer, DC numbers are increased and immature DCs decrease [41]. The allo-stimulatory capacity of DC and T cell proliferation against recall antigens was also enhanced.

Sorafenib, like sunitinib, inhibits receptor tyrosine kinases such as VEGFR-2 and VEGFR-3, PDGFR- β , Flt-3 and c-KIT [42]. However, it targets B-Raf [43] as well, which is a component of the Ras-Raf-MAPK pathway that controls T-cell activation [44]. It inhibits the activation of human peripheral blood T cells by targeting LCK phosphorylation [45]. In a recent publication sorafenib interferes with the function and maturation of the monocyte-derived DCs (MDDCs) by downregulating DC responsiveness to inflammatory signals [46]. DCs migration is reduced, as is their capacity for lymphocyte stimulation. In the same study sunitinib did not impair the induction of primary cell responses and reduced the number of T-regs.

Clinical efficacy and immune functions

Sunitinib has shown anticancer activity and has been approved for the use in first-line treatment of mRCC. It has a favorable side-effect profile, though in several clinical trials the development of side effects correlated with activity [47,48]. The correlation of clinical responses to immune induction may be an additional marker of ac-

tivity [37]. The positive effects of sunitinib on DCs functions are of major importance, as DCs are the most potent antigen-presenting cells and the orchestrators of the initial immune responses (Table 3). Whether the immunostimulatory effects of sunitinib account for the higher response rate seen in clinical trials or this represents just a pharmacodynamic readout remains to be answered.

Sorafenib displays different effects on the immune system. Additionally, it has shown lower response rates and could not establish a higher activity than IFN- α in first-line treatment. The sorafenib-mediated immune suppression may partially explain this phenomenon and could argue for trials to combine sorafenib with cytokine treatment [49]. A recent study, however, showed no benefit when low-dose IFN- α was added to sorafenib for first-line treatment in mRCC patients [50].

Finally, bevacizumab has shown benefit in the first-line treatment, in combination with IFN- α , whilst temsirolimus is effective alone and the addition of IFN- α results in inferior effectiveness. These two agents interfere with the DC physiology on a different way and this may explain the different results when combined with IFN- α . Table 2 resumes the effects of targeted therapies on the immune system of patients with advanced RCC.

New targeted treatments have changed the way we see and treat mRCC. There is still room to investigate the role of cytokines and the effects of the new agents on the immune system.

References

1. Jemal A, Siegel R, Ward E et al. Cancer statistics, 2008. *CA Cancer J Clin* 2008; 58: 71-96.
2. Beck SD, Patel MI, Snyder ME et al. Effect of papillary and chromophobe cell type on disease-free survival after nephrectomy for renal cell carcinoma. *Ann Surg Oncol* 2004; 11: 71-77.
3. Cheville JC, Lohse CM, Zincke H, Weaver AL, Blute ML. Comparisons of outcome and prognostic features among histologic subtypes of renal cell carcinoma. *Am J Surg Pathol* 2003; 27: 612-624.
4. Klimis T, Karvounis H. Renal cell carcinoma with rhabdoid features. Divergent differentiation of conventional (clear cell) carcinoma. *J BUON* 2008; 13: 433-436.
5. Motzer RJ, Mazumdar M, Bacik J, Berg W, Amsterdam A, Ferrara J. Survival and prognostic stratification of 670 pa-

- tients with advanced renal cell carcinoma. *J Clin Oncol* 1999; 17: 2530-2540.
6. Motzer RJ, Michaelson MD, Rosenberg J et al. Sunitinib efficacy against advanced renal cell carcinoma. *J Urol* 2007; 178: 1883-1887.
 7. Latif F, Tory K, Gnarr J et al. Identification of the von Hippel-Lindau disease tumor suppressor gene. *Science* 1993; 260: 1317-1320.
 8. George DJ, Kaelin WG Jr. The von Hippel-Lindau protein, vascular endothelial growth factor and kidney cancer. *N Engl J Med* 2003; 349: 419-421.
 9. Ferrara N, Kerbel R. Angiogenesis as a therapeutic target. *Nature* 2005; 438: 967-974.
 10. Guermonprez P, Valladeau J, Zitvogel L et al. Antigen presentation and T cell stimulation by dendritic cells. *Annu Rev Immunol* 2002; 20: 621-667.
 11. Chaux P, Moutet M, Martin M, Martin F. Tumor-infiltrating dendritic cells are defective in their antigen-presenting function and inducible B7 expression in rats. *Int J Cancer* 1997; 72: 619-624.
 12. Gabrilovich DI, Ciernik IF, Carbone DP. Dendritic cells in antitumor immune responses I. Defective antigen presentation in tumor-bearing hosts. *Cell Immunol* 1996; 170: 101-110.
 13. Ohm JE, Shurin MR, Esche C et al. Effect of vascular endothelial growth factor and FLT3 ligand on dendritic cell generation in vivo. *J Immunol* 1999; 163: 3260-3268.
 14. Gabrilovich DI, Chen HL, Girgis KR et al. Production of vascular endothelial growth factor by human tumors inhibits the functional maturation of dendritic cells. *Nat Med* 1996; 2: 1096-1103.
 15. Takahashi A, Kono K, Ichihara F et al. Vascular endothelial growth factor inhibits maturation of dendritic cells induced by lipopolysaccharide, but not by proinflammatory cytokines. *Cancer Immunol Immunother* 2004; 53: 543-550.
 16. Dikov MM, Ohm JE, Ray N et al. Differential roles of vascular endothelial growth factor receptors 1 and 2 in dendritic cell differentiation. *J Immunol* 2005; 174: 215-222.
 17. Fricke I, Mirza N, Dupont J et al. Vascular endothelial growth factor-trap overcomes defects in dendritic cell differentiation but does not improve antigen-specific immune responses. *Clin Cancer Res* 2007; 13: 4840-4848.
 18. Ohm JE, Gabrilovich DI, Sempowski GD et al. VEGF inhibits T-cell development and may contribute to tumor-induced immune suppression. *Blood* 2003; 101: 4878-4886.
 19. Negrier S, Escudier B, Lasset C et al. Recombinant human IL-2, recombinant human interferon alpha-2a, or both in metastatic renal-cell carcinoma. *Groupe Francais d'Immunotherapie. N Engl J Med* 1998; 338: 1272-1278.
 20. Motzer RJ, Russo P. Systemic therapy for renal cell carcinoma. *J Urol* 2000; 163: 408-417.
 21. Negrier S, Perol D, Ravaud A et al. Medroxyprogesterone, interferon alfa-2a, interleukin 2, or combination of both cytokines in patients with metastatic renal carcinoma of intermediate prognosis: results of a randomized controlled trial. *Cancer* 2007; 110: 2468-2477.
 22. Radulovic S, Bjelogrić SK. Sunitinib, sorafenib and mTOR inhibitors in renal cancer. *J BUON* 2007; 12 (Suppl 1): S151-S162.
 23. Motzer RJ, Rini BI, Bukowski RM et al. Sunitinib in patients with metastatic renal cell carcinoma. *JAMA* 2006; 295: 2516-2524.
 24. Motzer RJ, Hutson TE, Tomczak P et al. Sunitinib versus interferon alpha in metastatic renal-cell carcinoma. *N Engl J Med* 2007; 356: 115-124.
 25. Kontovinis LF, Papazisis KT, Touplikioti P et al. Sunitinib treatment for patients with clear-cell metastatic renal cell carcinoma: clinical outcomes and plasma angiogenesis markers. *BMC Cancer* 2009; 9: 82.
 26. Escudier B, Eisen T, Stadler WM et al. Sorafenib in advanced clear-cell renal-cell carcinoma. *N Engl J Med* 2007; 356: 115-134.
 27. Escudier B, Pluzanska A, Koralewski P et al. Bevacizumab plus interferon alfa-2a for treatment of metastatic renal cell carcinoma: a randomised, double-blind phase III trial. *Lancet* 2007; 370: 2103-2111.
 28. Hudes G, Carducci M, Tomczak P et al. Temsirolimus, interferon alfa, or both for advanced renal-cell carcinoma. *N Engl J Med* 2007; 356: 2271-2281.
 29. Motzer RJ, Escudier B, Oudard S et al. Efficacy of everolimus in advanced renal cell carcinoma: a double-blind, randomized, placebo-controlled phase III trial. *Lancet* 2008; 372: 449-456.
 30. Gabrilovich DI, Ishida T, Nadaf S et al. Antibodies to vascular endothelial growth factor enhance the efficacy of cancer immunotherapy by improving endogenous dendritic cell function. *Clin Cancer Res* 1999; 5: 2963-2970.
 31. Tamir A, Jordan WJ, Ritter M et al. Interferon-alpha2a is sufficient for promoting dendritic cell immunogenicity. *Clin Exp Immunol* 2005; 142: 471-480.
 32. Melichar B, Koralewski P, Ravaud A et al. First-line bevacizumab combined with reduced dose interferon-alpha2a is active in patients with metastatic renal cell carcinoma. *Ann Oncol* 2008; 19: 1470-1476.
 33. Cao W, Manicassamy S, Tang H et al. Toll-like receptor-mediated induction of type I interferon in plasmacytoid dendritic cells requires the rapamycin-sensitive PI(3)K-mTOR-p70S6K pathway. *Nat Immunol* 2008; 9: 1157-1164.
 34. Ko H, Hambly BD, Eris JM et al. Dendritic cell derived IL-18 production is inhibited by rapamycin and sanglifehrin A, but not cyclosporine A. *Transpl Immunol* 2008; 20: 99-105.
 35. Woltman AM, van der Kooij SW, Coffey PJ et al. Rapamycin specifically interferes with GM-CSF signaling in human dendritic cells, leading to apoptosis via increased p27KIP1 expression. *Blood* 2003; 101: 1439-1445.
 36. Ohtani M, Nagai S, Kondo S et al. Mammalian target of rapamycin and glycogen synthase kinase 3 differentially regulate lipopolysaccharide-induced interleukin-12 production in dendritic cells. *Blood* 2008; 112: 635-643.
 37. Weichhart T, Costantino G, Poglitsch M et al. The TSC-mTOR signaling pathway regulates the innate inflammatory response. *Immunity* 2008; 29: 565-577.
 38. Van Crujisen H, van der Veldt AA, Vroeling L et al. Sunitinib-induced myeloid lineage redistribution in renal cell cancer patients: CD1c+ dendritic cell frequency predicts progression-free survival. *Clin Cancer Res* 2008; 14: 5884-5892.
 39. Lenahan C, Cho D, Bissonnette A et al. Immunologic effects of sunitinib in renal cell carcinoma. *J Clin Oncol* 2008; 26 (Suppl): 14551 (abstr).
 40. Finke J, Rini B, Ireland J, Rayman P et al. Sunitinib reverses type-I immune suppression and decreases T-regulatory cells in renal cell carcinoma patients. *Clin Cancer Res* 2008; 14: 6674-6682.
 41. Osada T, Chong G, Tansik R et al. The effect of anti-VEGF therapy on immature myeloid cell and dendritic cells in cancer patients. *Cancer Immunol Immunother* 2008; 57: 1115-1124.
 42. Wilhelm S, Chien DS. BAY 43-9006: preclinical data. *Curr Pharm Des* 2002; 8: 2255-2257.

43. Wilhelm SM, Carter C, Tang L et al. BAY 43-9006 exhibits broad spectrum oral antitumor activity and targets the RAF/MEK/ERK pathway and receptor tyrosine kinases involved in tumor progression and angiogenesis. *Cancer Res* 2004; 64: 7099-7109.
44. Franklin RA, Tordai A, Patel H et al. Ligation of the T cell receptor complex results in activation of the Ras/Raf-1/MEK/MAPK cascade in human T lymphocytes. *J Clin Invest* 1994; 93: 2134-2140.
45. Zhao W, Gu YH, Song R et al. Sorafenib inhibits activation of human peripheral blood T cells by targeting LCK phosphorylation. *Leukemia* 2008; 22: 1226-1233.
46. Hipp MM, Hiff N, Walter S et al. Sorafenib, but not sunitinib, affects function of dendritic cells and induction of primary immune responses. *Blood* 2008; 111: 5610-5620.
47. Rixe O, Billemonet B, Izzedine H. Hypertension as a predictive factor of sunitinib activity. *Ann Oncol* 2007; 18: 1117.
48. Wolter P, Stefan C, Decallonne B et al. Evaluation of thyroid dysfunction as a candidate surrogate marker for efficacy of sunitinib in patients (pts) with advanced renal cell carcinoma (RCC). *J Clin Oncol* 2008; 26 (Suppl): 5126 (abstr).
49. Eto M, Takeuchi A, Ohki T et al. In vitro and in vivo analysis of synergistic antitumor effects of interferon- α and sorafenib in renal cell carcinoma. *J Clin Oncol* 2008; 26 (Suppl): 16143 (abstr).
50. Tannir NM, Zurita AJ, Heymach JV et al. A randomized phase II trial of sorafenib versus sorafenib plus low-dose interferon- α : Clinical results and biomarker analysis. *J Clin Oncol* 2008; 26(Suppl): 5093 (abstr).
51. Rini BI. Metastatic renal cell carcinoma: many treatment options, one patient. *J Clin Oncol* 2009; 27: 3225-3234.
52. Fyfe G, Fisher RI, Rosenberg SA et al. Results of treatment of 255 patients with metastatic renal cell carcinoma who received high-dose recombinant interleukin-2 therapy. *J Clin Oncol* 1995; 13: 688-696.
53. Klapper JA, Downey SG, Smith FO et al. High-dose interleukin-2 for the treatment of metastatic renal cell carcinoma: a retrospective analysis of response and survival in patients treated in the surgery branch at the National Cancer Institute between 1986 and 2006. *Cancer* 2008; 113: 293-301.
54. McDermott DF, Regan MM, Clark JI et al. Randomized phase III trial of high-dose interleukin-2 versus subcutaneous interleukin-2 and interferon in patients with metastatic renal cell carcinoma. *J Clin Oncol* 2005; 23: 133-141.
55. Yang JC, Sherry RM, Steinberg SM et al. Randomized study of high-dose and low-dose interleukin-2 in patients with metastatic renal cancer. *J Clin Oncol* 2003; 21: 3127-3132.
56. Interferon- α and survival in metastatic renal carcinoma: early results of a randomized controlled trial: Medical Research Council, Renal Cancer Collaborators. *Lancet* 1999; 353: 14-17.
57. Coppin C, Porzolt F, Awa A et al. Immunotherapy for advanced renal cell cancer. *Cochrane Database Syst Rev*: CD001425, 2005.
58. Bukowski RM. Cytokine therapy for metastatic renal cell carcinoma. *Semin Urol Oncol* 2001; 19: 148-154.
59. Motzer RJ, Bacik J, Murphy BA et al. Interferon- α as a comparative treatment for clinical trials of new therapies against advanced renal cell carcinoma. *J Clin Oncol* 2002; 20: 289-296.
60. Motzer RJ, Michaelson MD, Redman BG et al. Activity of SU11248, a multitargeted inhibitor of vascular endothelial growth factor receptor and platelet-derived growth factor receptor, in patients with metastatic renal cell carcinoma. *J Clin Oncol* 2006; 24: 16-24.
61. Szczylik C, Demkow T, Staehler M et al. Randomized phase III trial of first-line treatment with sorafenib versus interferon in patients with advanced renal cell carcinoma: final results. *J Clin Oncol* 2007; 25: 5025.
62. Figlin RA, Hutson TE, Tomczak P et al. Overall survival with sunitinib versus interferon (IFN)- α as first-line treatment of metastatic renal cell carcinoma (MRCC). *J Clin Oncol* 2008; 26(Suppl): 256S (abstr 5024).
63. Bukowski RM, Eisen T, Szczylik C et al. Final results of the randomized phase III trial of sorafenib in advanced renal cell carcinoma: survival and biomarker analysis. *J Clin Oncol* 2007; 25(Suppl): 240S (abstr 5023).
64. Yang JC, Haworth L, Sherry RM et al. Randomized trial of bevacizumab, an anti-vascular endothelial growth factor antibody, for metastatic renal cancer. *N Engl J Med* 2003; 349: 427-434.
65. Bukowski RM, Kabbinavar FF, Figlin RA et al. Randomized phase III study of erlotinib combined with bevacizumab compared with bevacizumab alone in metastatic renal cell cancer. *J Clin Oncol* 2007; 25: 4536-4541.
66. Atkins MB, Hidalgo M, Stadler WM et al. Randomized phase II study of multiple dose levels of CCI-779, a novel mammalian target of rapamycin kinase inhibitor, in patients with advanced refractory renal cell carcinoma. *J Clin Oncol* 2004; 22: 909-912.