

ORIGINAL ARTICLE

Novel insights into laryngeal squamous cell carcinoma from association study of aberrantly expressed miRNAs, lncRNAs and clinical features in Bulgarian patients

Silva Garo Kyurkchiyan¹, Todor Miroslavov Popov², Gergana Stancheva¹, Julian Rangachev², Vanyo Ivanov Mitev¹, Diana Petrova Popova², Radka Petrova Kaneva¹

¹Molecular Medicine Center, Department of Medical Chemistry and Biochemistry, Medical University, Sofia, Bulgaria; ²Clinic of ENT, Department of ENT, Medical Faculty, Medical University, Sofia, Bulgaria.

Summary

Purpose: Laryngeal cancer is one of most common and aggressive head and neck cancers with poor prognosis and great necessity for improvement of treatment modalities. MicroRNAs (miRs) are among the most investigated molecules recently due to their potential as diagnostic and prognostic biomarkers in cancer. The purpose of our study was to explore the association of certain clinicopathological features with the expression levels of some known cancer associated non-coding (nc) RNAs: miR-21 and miR-31 in both of their isoforms, miR-145-5p, miR-55-5p, miR-196a-5p, miR-210-3p, miR-221-3p, miR-222-3p, miR-424-5p, lncRNA MALAT1 and lncRNA HOTAIR.

Methods: Expression levels of the chosen markers were investigated in laryngeal squamous cell carcinoma (LSCC) and normal samples in 82 Bulgarian patients via RT-qPCR, and the results were analyzed with SPSS v23.0 statistical software.

Results: All of the explored ncRNAs were significantly downregulated in LSCC samples, suggesting their involvement in laryngeal carcinogenesis. New significant association were found between the expression levels of miR-21-5p, miR-222-3p, HOTAIR and family history. Moreover, miR-424-5p showed potential as marker for subglottic LSCC location, and "passenger" miR-31-3p was significantly upregulated in well and moderately differentiated LSCC.

Conclusion: Our results enrich the knowledge about ncRNA involvement in LSCC tumorigenesis. Further studies are needed to evaluate the clinical utility of the differently expressed ncRNAs as potential diagnostic and prognostic biomarkers in LSCC.

Key words: expression, laryngeal cancer, lncRNAs, miRNAs, novel insights

Introduction

Worldwide, cancer of the larynx is the eleventh most common of all malignant neoplasms in men (GLOBOCAN 2012, IACR) [1]. According to the National Cancer Registry of Bulgaria for 2015, 554 (3.1%) patients were diagnosed and 343 (3.3%) were the registered deaths from laryngeal neoplasm [2], which places Bulgaria on the 9th place of laryngeal cancer incidence and mortality in Europe, with 8.9 incidence cases per 100 000 (GLOBOCAN 2018, IACR) [3]. This data shows that Bulgaria has one

of the highest morbidity rates in Europe, indicating the social significance of this disease. The clinical features depending on the laryngeal tumour location may vary very early, which is a prerequisite for early diagnosis and good prognosis, but still laryngeal cancer often is diagnosed in advanced stage (III or IV stage) [4], as patients neglect their symptoms. Despite modern medical techniques of cancer surgery and treatment, laryngeal cancer is still a challenging disease with not significantly

Corresponding author: Silva Garo Kyurkchiyan, MSc. Molecular Medicine Center, Department of Medical Chemistry and Biochemistry, Medical University - Sofia, 2 Zdrave Str., 1431, Sofia, Bulgaria.
Tel: +359887762467, Email: sggiragosyan@abv.bg
Received: 27/05/2019; Accepted: 02/08/2019

changed 5-year survival for decades, which account about 60% of diagnosed patients [5]. Novel molecular genetic markers are required in order to improve clinical LSCC diagnosis and prognosis.

With the discovery of the oncogenes and tumour suppressors involvement in long non-coding RNAs (lncRNAs) [6] and miRs [7] in the early 2000s, it was postulated their potential role as biomarkers due to their deregulation associated with various cellular processes in neoplasms [8]. MiRs are responsible for the expression of two mature miRs: one from the 5' strand and one from the 3' strand of the precursor (miR-5p and miR-3p). However, one of the isoforms, called the "guide" strand, is usually much more prevalent and more biologically active than the other isoform, the "passenger" strand, which is known as miR* [9]. Changes in the expression of these RNA molecules contribute to the formation and progression of cancer and have a key role in tumour microenvironment [10]. Classic example is the article of Medina et al [11] in 2010, which proves that overexpression of only one (miR-21) is enough to give rise to malignant tumours. Additionally, when inhibited, the same tumour lesions regress - i.e. miR tumour "dependency" was first described [11].

However, in people, the latest release of the miR database (miRBase) has catalogued 1 917 precursors and 2 654 miRNAs (miRBase release 22.1, update October 2018) [12], whereas the number of annotated lncRNAs in NONCODE database (v5) is 172 216 transcripts, coded by 96 308 genes [13]. Despite the great number of research projects, exploring the clinical significance of these ncRNAs, the functional importance of many of those remains to be determined. Understanding the non-coding RNA world, which represents a gold mine for novel biomedical markers and therapeutic strategies, is one of the most important challenges in front of the molecular biology and biomedicine.

The aim of this study was to explore further the role of ncRNAs and their potential as biomarkers in LSCC. We investigated the expression levels of 13 ncRNAs known to be deregulated ncRNAs in cancers, chosen from the literature database searching (PubMed, Scholar, etc.) in laryngeal specimens. Eleven miRs isoforms: miR-21, miR-31, miR-145-5p, miR-155-5p, miR-196a-5p, miR-210-3p, both miR-221-3p/222-3p and miR-424-5p, and two lncRNAs: lncRNA MALAT1 (Metastasis-associated lung adenocarcinoma transcript 1) and lncRNA HOTAIR (Hox antisense intergenic RNA). MiR-21 and miR-31 were investigated in both isoforms (3p and 5p) due to lack of sufficient knowledge about the function of "passenger" isoforms: miR-21-3p and miR-31-3p.

Methods

Patients and tissue samples

Fresh-frozen tumour and normal tissue was obtained from a total of 82 patients diagnosed with LSCC and enrolled in the current study. Patients were recruited at the Ear, Nose and Throat Department, University Hospital "Queen Joanna - ISUL" Sofia during 2012-2016. Informed consent was obtained from each patient, and the current study was approved by the Ethics Committee of Medical University of Sofia with protocols no. 13/23.04.2015, no./22.04.2016, no.432/2017 and no.435/2017. Samples were stored at -80°C until use at the Molecular Medicine Center biobank at the Department of Medical Chemistry and Biochemistry, Medical University - Sofia. Additional material from formalin-fixed, paraffin-embedded (FFPE) tissue was used to conduct immunohistochemical analysis to determine the clinicopathological characteristics of the samples. None of the patients in this study had received chemotherapy or radiotherapy before surgery.

Isolation of total RNA from tissue material and reverse transcription

Total RNA was isolated from fresh frozen tumour and normal tissue using RNA extraction kit from tissue (miRNeasy Micro Kit, Qiagen, Hilden, Germany). The quality and quantity of the total RNA samples were assessed by denaturing electrophoresis on a formaldehyde gel and NanoDrop 2000 (ThermoFisher Scientific, Waltham, DE, USA).

500 ng total RNA of each sample were used to prepare cDNA by using miScript II RT Kit and RT² First Strand Kit (Qiagen, Hilden, Germany), according to the manufacturer's protocol. Afterward, reverse transcription (RT) samples were processed further or stored at -20°C until use.

Real-time quantitative polymerase chain reaction (RT-qPCR)

The expression of mature miRs and lncRNAs was assayed using miScript SYBR Green PCR kit and RT² SYBR Green Mastermix (Qiagen, Hilden, Germany) on a 7900HT Fast Real Time PCR System (Applied Biosystems, California, USA). The miScript Primer assays (Qiagen, Hilden, Germany) were used for 11 mature miRs: miR-21-3p, miR-21-5p, miR-31-3p, miR-31-5p, miR-145-5p, miR-155-5p, miR-196a-5p, miR-210-3p, miR-221-3p, miR-222-3p and miR-424-5p. The RT² lncRNA qPCR assays (Qiagen, Hilden, Germany) were used for 2 long non-coding RNAs: lncRNA MALAT1 and lncRNA HOTAIR. Each reaction was performed in triplicate in a total volume of 10 µL, according to the manufacturer's protocol. Expression levels of miRs were normalized to the internal control RNU6-2 (Qiagen, Hilden, Germany) and for lncRNAs Actin mRNA (Qiagen, Hilden, Germany) was used as control. The relative quantification (RQ) of miRs and lncRNAs in tumour samples was analyzed by the 2- $\Delta\Delta C_t$ method, as previously described [14]. A RQ ≥ 2 was defined as overexpression, RQ < 0.5 as underexpression, and RQ between 1.99 and 0.5 as no change in expression.

Statistics

Data analysis was performed with the SPSS software version 23.0 for Windows (IBM SPSS, NY, USA). The expression levels of the studied miRs were evaluated in 82 laryngeal tumour and adjacent normal laryngeal squamous cell tissue specimens, whereas lncRNA MALAT1 and lncRNA HOTAIR were evaluated in 63 tumour and adjacent normal laryngeal samples. Kolmogorov-Smirnov test for normality, Wilcoxon test, Mann-Whitney U test, Kruskal-Wallis or one-way ANOVA test, paired and unpaired T-tests were used as appropriate. A value of $p < 0.05$ was considered statistically significant.

Results

In the current study, 4 females and 78 males, in total 82 LSCC patients, were included. The mean age of the patient group was 61 years (range 41-84). Clinicopathological characteristics of the patients are shown in Tables 1a and 1b together with the expression levels of the explored ncRNAs.

Relative expression in laryngeal tumour samples

All of the investigated ncRNAs in LSCC tissue samples were significantly differentially expressed in tumour tissue in comparison to paired normal tissue. However, the relative expression levels of miR-21-3p ($p < 0.001$), miR-21-5p ($p < 0.001$), miR-31-3p ($p = 0.004$), miR-31-5p ($p < 0.001$), miR-155-5p ($p = 0.003$), miR-210-3p ($p = 0.025$), miR-221-3p

($p = 0.001$), miR-222-3p ($p = 0.001$), miR-196a-5p ($p < 0.001$) and lncRNA HOTAIR ($p = 0.004$) were significantly upregulated, whereas miR-145-5p ($p < 0.001$) and lncRNA MALAT1 ($p < 0.001$) were significantly downregulated in laryngeal tumour tissue in comparison to normal tissue. Reversed and normalized dCt values are shown in boxplots (Figure 1). Depending on the normality distribution tests of the samples, Wilcoxon test or paired T-test were used to evaluate the level of significance of the expression differences between tumour and normal laryngeal tissue.

Association between ncRNA expression levels and clinicopathological characteristics

Association between RQ data of the studied ncRNAs and the groups based on the clinicopathological characteristics of the LSCC patients is summarized in Tables 1a and 1b. Our data indicates statistically significant association between the expression levels of miR-21-5p, miR-31-3p, miR-31-5p, miR-145-5p, miR-222-3p, miR-424-5p, lncRNA MALAT1, lncRNA HOTAIR and different clinical features. MiR-21-5p showed statistically significant association with tumour differentiation ($p = 0.036$), positive family history ($p = 0.050$) and work exposures ($p = 0.046$). MiR-31-3p was associated with positive nodal metastasis ($p = 0.009$) and tumour differentiation ($p = 0.034$). The expression

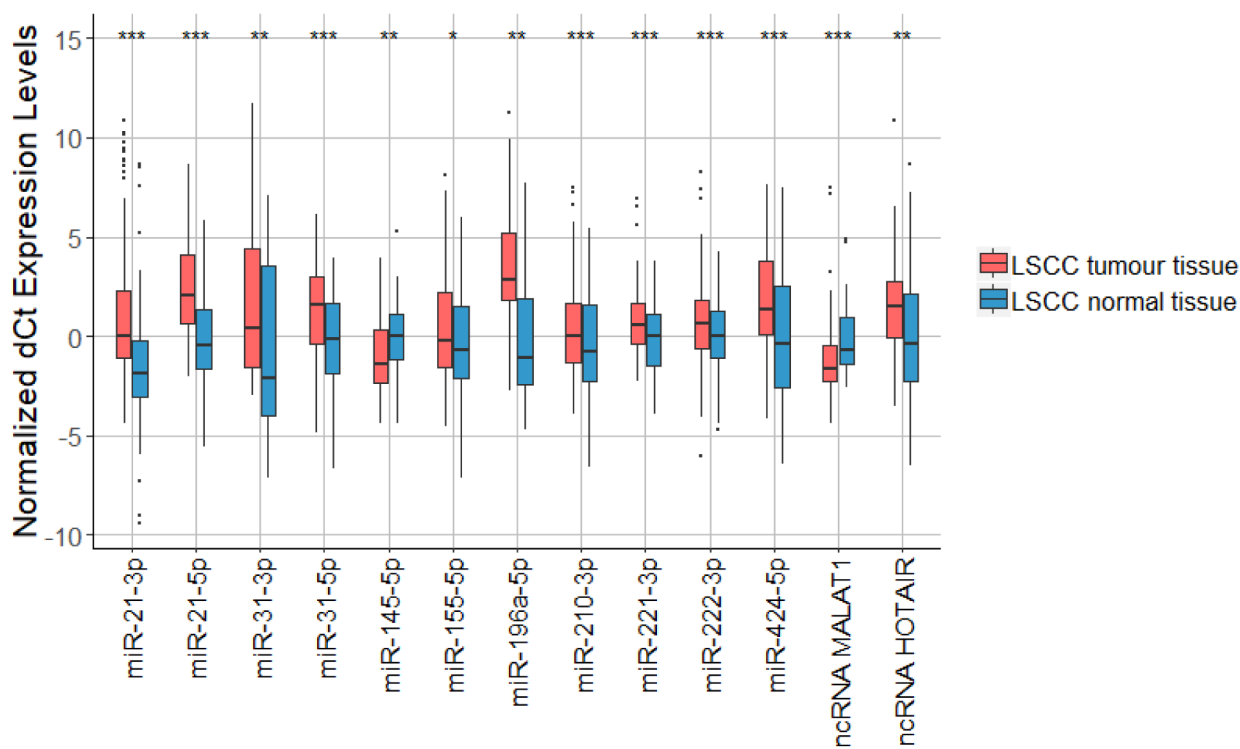


Figure 1. Normalized dCt expression levels of investigated ncRNAs in laryngeal tumour and normal tissue with levels of significance: * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.

Table 1a. Association between mean RQ levels of miR-21 and miR-31 isoforms, miR-145-5p, miR-155-5p, miR-196a-5p and clinicopathological characteristics in patients with LSCC

Clinical features	Number of patients (%)	Relative expression of miR-21-3p (mean±SD)	Relative expression of miR-21-5p (mean±SD)	Relative expression of miR-31-3p (mean±SD)	Relative expression of miR-31-5p (mean±SD)	Relative expression of miR-145-5p (mean±SD)	Relative expression of miR-155-5p (mean±SD)	Relative expression of miR-196a-5p (mean±SD)
Gender								
Female	4 (7.88)	p=0.666 5.000 ± 0.169	p=0.613 6.055 ± 2.863	p=0.754 6.750±8.131	p=0.797 6.825±8.167	p=0.178 0.248 ± 0.061	p=0.393 0.946 ± 0.964	p=0.314 48.606±67.352
Male	78 (95.12)	6.468 ± 6.345	8.166 ± 9.434	7.390±13.956	6.939±10.384	1.020 ± 1.331	4.478 ± 6.353	32.979±39.498
Age, years								
≤60	49 (59.76)	p=0.446 6.926 ± 5.161	p=0.902 8.575 ± 9.131	p=0.380 7.950±10.949	p=0.305 8.771±11.134	p=0.738 0.815 ± 0.879	p=0.313 5.925 ± 7.382	p=0.314 42.274±47.878
>60	33 (40.24)	6.104 ± 6.908	8.372±10.610	6.432±13.117	6.202±10.153	0.563 ± 0.521	3.097 ± 4.264	27.731±33.167
Tumour stage								
T1/2	11 (13.41)	p=0.301 4.065 ± 2.597	p=0.832 7.284 ± 4.976	p=0.985 9.518±19.366	p=0.048 2.007±1.870	p=0.035 1.253 ± 1.408	p=0.642 3.957 ± 4.092	p=0.746 23.635±38.040
T3	28 (34.15)	7.902 ± 6.481	8.681±11.655	6.303±8.884	6.643±9.745	0.741 ± 0.684	6.390 ± 8.151	40.030±46.452
T4	43 (52.45)	5.976 ± 6.579	7.482 ± 9.518	7.589±15.376	9.443±8.121	0.459 ± 0.319	2.752 ± 3.410	29.967±39.456
Nodal stage								
N0 negative	56 (68.29)	p=0.957 6.294 ± 5.514	p=0.834 8.238 ± 7.618	p=0.009 8.749±15.585	p=0.805 6.858±10.090	p=0.007 1.102 ± 1.315	p=0.948 4.645 ± 6.233	p=0.597 31.126±39.661
N1-3 positive	26 (30.71)	6.838 ± 8.209	6.441 ± 4.603	3.319±3.568	8.318±12.071	0.485 ± 0.502	3.006 ± 4.585	40.639±41.387
Tumour differentiation								
G1 well differentiation	31 (37.80)	p=0.429 4.410 ± 4.326	p=0.036 7.421 ± 8.011	p=0.034 8.553±19.122	p=0.797 7.163±10.175	p=0.722 0.790 ± 0.789	p=0.374 4.066 ± 5.130	p=0.219 26.399±35.983
G2 moderate differentiation	40 (48.78)	8.233 ± 7.365	7.352 ± 6.491	7.314±9.642	7.602±11.497	0.725 ± 0.781	4.465 ± 6.668	40.863±43.937
G3 poor differentiation	11 (13.41)	4.332 ± 3.471	15.480±8.724	1.535±2.209	4.791±4.346	0.232 ± 0.158	3.300 ± 1.863	16.216±10.771
Tumour localization								
Supraglottis	39 (47.56)	p=0.259 6.361 ± 7.414	p=0.376 7.375 ± 8.064	p=0.288 4.205±4.257	p=0.516 5.327±7.014	p=0.288 0.423 ± 0.418	p=0.821 2.986 ± 4.554	p=0.641 33.632±39.336
Glottis	45 (54.87)	6.437 ± 6.284	7.686 ± 6.717	9.061±16.736	8.250±11.889	0.819 ± 0.781	5.495 ± 6.750	35.248±44.407
Subglottis	8 (9.75)	6.590 ± 2.278	9.181 ± 6.050	7.544±14.361	7.334±12.024	0.957 ± 1.153	1.636 ± 1.299	25.816±16.863
Family history								
Deny	63 (60.97)	p=0.370 6.399 ± 6.681	p=0.050 5.354 ± 3.852	p=0.653 7.804±14.941	p=0.721 6.765±10.574	p=0.141 0.715 ± 0.696	p=0.705 3.675 ± 4.450	p=0.987 34.281±42.603
Report	19 (23.18)	6.532 ± 4.880	9.167 ± 8.046	6.085 ± 9.721	8.591±10.675	0.711 ± 0.957	5.849 ± 8.637	31.400±32.158
Missing data	13 (15.85)							
Tobacco smoking								
Deny	4 (4.87)	p=0.744 5.700 ± 2.880	p=0.205 3.910 ± 3.631	p=0.516 3.166 ± 3.085	p=0.687 4.673 ± 6.161	p=0.048 1.450 ± 1.753	p=0.678 2.431 ± 1.979	p=0.920 12.344±13.935
≤20 cigarettes per day	26 (31.70)	7.316 ± 6.702	10.148±7.889	9.599±17.740	6.543±5.778	0.950 ± 0.913	4.846 ± 5.412	40.942±49.079
21 < ≥40 cigarettes per day	30 (36.58)	5.866 ± 5.075	6.522 ± 4.110	5.356±10.160	9.361±15.564	0.653 ± 0.435	2.851 ± 3.694	38.790±43.680
≥41 cigarettes per day	13 (15.85)	7.007 ± 8.722	7.604 ± 9.332	4.294 ± 3.197	7.144±11.791	0.318 ± 0.291	5.408 ± 8.979	20.553±22.720
Missing data	9 (10.97)							
Alcohol consumption								
Deny	13 (15.85)	p=0.600 5.062 ± 3.224	p=0.768 6.916 ± 4.696	p=0.714 6.332±12.692	p=0.754 6.922±10.650	p=0.295 0.758 ± 1.054	p=0.278 4.775 ± 9.738	p=0.670 16.235±16.313
<100 mL per day	20 (24.39)	7.173 ± 5.025	10.280±9.674	7.622±10.690	6.052 ± 5.113	0.840 ± 0.962	3.678 ± 5.049	41.079±46.826
100 - 200 mL per day	19 (23.17)	5.697 ± 4.826	8.944 ± 7.814	9.117±19.506	8.686±14.673	0.627 ± 0.518	3.543 ± 4.194	34.454±52.613
>200 mL per day	17 (20.73)	6.727 ± 7.886	6.548 ± 5.371	4.190 ± 4.886	9.866±12.675	0.811 ± 0.793	5.571 ± 5.112	38.359±33.988
Missing data	13 (15.85)							
Work exposures								
Deny	46 (40.25)	p=0.258 7.283 ± 7.225	p=0.046 5.594 ± 3.711	p=0.081 5.818 ± 3.911	p=0.794 7.827±11.751	p=0.666 0.827 ± 1.154	p=0.863 3.468 ± 4.022	p=0.481 27.671±32.926
Report	36 (43.90)	5.334 ± 4.558	10.473±7.386	8.404±13.510	6.458 ± 8.897	1.110 ± 1.307	5.210 ± 7.534	41.138±47.161
Missing data	13 (15.85)							

Table 1b. Association between mean RQ levels of miR-210-3p, miR-221-3p, miR-222-3p, miR-424-5p, lncRNA MALAT1, lncRNA HOTAIR and clinicopathological characteristics in patients with LSCC

Clinical features	Number of patients (%)	Relative expression of miR-210-3p (mean±SD)	Relative expression of miR-221-3p (mean±SD)	Relative expression of miR-222-3p (mean±SD)	Relative expression of miR-424-5p (mean±SD)	Relative expression of lncRNA MALAT1 (mean±SD)	Relative expression of lncRNA HOTAIR (mean±SD)
Gender							
Female	4 (7.88)	p=0.386 2.379 ± 0.585	p=0.829 1.926 ± 0.807	p=0.023 1.566 ± 0.828	p=0.795 6.731 ± 1.595	p=0.077 0.209 ± 0.113	p=0.060 2.529 ± 3.507
Male	78 (95.12)	p=0.574 3.167 ± 3.975	p=0.817 2.691 ± 3.036	p=0.052 3.502 ± 5.577	p=0.817 9.906 ± 12.037	p=0.024 0.739 ± 0.986	p=0.130 6.472 ± 8.638
Age, years							
≤60	49 (59.76)	2.369 ± 2.959	2.541 ± 2.629	3.189 ± 3.970	10.071 ± 10.851	1.076 ± 1.008	7.674 ± 9.580
>60	33 (40.24)	3.651 ± 4.394	2.878 ± 3.208	5.035 ± 4.118	9.604 ± 12.601	0.634 ± 1.123	5.432 ± 7.762
Tumour stage							
T1/2	11 (13.41)	p=0.383 1.586 ± 1.650	p=0.144 2.002 ± 1.058	p=0.857 1.701 ± 1.711	p=0.873 6.626 ± 4.683	p=0.072 0.431 ± 0.648	p=0.918 3.576 ± 4.429
T3	28 (34.15)	3.031 ± 3.872	1.477 ± 0.877	4.798 ± 5.265	11.382 ± 11.620	0.975 ± 1.381	6.437 ± 8.644
T4	43 (52.45)	3.603 ± 4.299	5.002 ± 7.985	2.890 ± 3.763	9.446 ± 13.249	0.514 ± 0.442	6.940 ± 9.271
Nodal stage							
N0 negative	56 (68.29)	p=0.562 3.128 ± 3.865	p=0.743 3.281 ± 5.744	p=0.597 2.456 ± 6.062	p=0.012 9.288 ± 13.175	p=0.915 0.696 ± 1.043	p=0.392 5.582 ± 7.271
N1-3 positive	26 (30.71)	3.166 ± 4.169	4.026 ± 8.231	3.361 ± 4.757	4.733 ± 3.663	0.598 ± 0.507	8.515 ± 11.493
Tumour differentiation							
G1 well differentiation	31 (37.80)	p=0.795 2.989 ± 3.131	p=0.126 5.251 ± 9.919	p=0.139 4.618 ± 5.429	p=0.689 7.333 ± 10.337	p=0.481 0.558 ± 0.514	p=0.243 4.028 ± 5.220
G2 moderate differentiation	40 (48.78)	3.555 ± 4.601	2.303 ± 2.078	2.672 ± 3.158	7.787 ± 8.934	0.790 ± 1.179	7.801 ± 10.093
G3 poor differentiation	11 (13.41)	3.076 ± 1.836	2.879 ± 1.811	2.905 ± 1.447	9.901 ± 16.039	0.371 ± 0.288	7.366 ± 8.989
Tumour localization							
Supraglottis	39 (47.56)	p=0.886 3.141 ± 3.956	p=0.702 1.961 ± 1.734	p=0.293 1.796 ± 1.273	p=0.015 4.216 ± 4.125	p=0.530 0.648 ± 0.457	p=0.417 9.265 ± 10.490
Glottis	45 (54.87)	2.873 ± 3.385	4.577 ± 8.278	4.729 ± 5.327	9.661 ± 13.161	0.746 ± 1.180	3.978 ± 5.034
Subglottis	8 (9.75)	4.403 ± 6.023	2.240 ± 1.578	1.659 ± 1.182	12.023 ± 6.780	0.393 ± 0.434	9.610 ± 12.960
Family history							
Deny	63 (60.97)	p=0.952 3.117 ± 3.850	p=0.917 3.181 ± 5.769	p=0.015 2.567 ± 4.022	p=0.987 9.541 ± 11.922	p=0.868 0.703 ± 1.027	p=0.031 4.748 ± 5.678
Report	19 (23.18)	3.201 ± 4.209	4.319 ± 8.139	4.754 ± 5.408	10.522 ± 11.963	0.577 ± 0.593	10.145 ± 11.211
Missing data	13 (15.85)						
Tobacco smoking							
Deny	4 (4.87)	p=0.453 6.894 ± 8.849	p=0.279 3.295 ± 1.822	p=0.499 2.282 ± 1.605	p=0.174 10.673 ± 9.034	p=0.086 0.123 ± 0.087	p=0.275 4.606 ± 6.775
≤20 cigarettes per day	26 (31.70)	3.567 ± 4.494	4.302 ± 7.917	4.297 ± 5.803	14.022 ± 16.123	0.939 ± 1.419	5.515 ± 9.937
21 < ≥40 cigarettes per day	30 (36.58)	2.502 ± 2.600	2.329 ± 2.162	2.947 ± 4.104	8.976 ± 9.986	0.653 ± 0.532	7.411 ± 7.146
≥41 cigarettes per day	13 (15.85)	3.151 ± 3.214	4.525 ± 9.297	3.725 ± 5.382	4.888 ± 5.974	0.395 ± 0.255	4.005 ± 4.067
Missing data	9 (10.97)						
Alcohol consumption							
Deny	13 (15.85)	p=0.146 2.990 ± 5.342	p=0.154 2.046 ± 1.383	p=0.506 2.257 ± 1.679	p=0.892 9.990 ± 6.572	p=0.108 0.758 ± 1.054	p=0.391 5.317 ± 6.982
<100 mL per day	20 (24.39)	2.590 ± 1.912	5.295 ± 10.256	4.674 ± 4.949	8.335 ± 8.433	0.946 ± 0.997	9.981 ± 12.399
from 100 to 200 mL per day	19 (23.17)	3.484 ± 3.992	2.634 ± 2.473	3.532 ± 4.810	11.953 ± 14.090	1.067 ± 1.356	3.469 ± 4.847
>200 mL per day	17 (20.73)	4.272 ± 4.749	5.222 ± 9.002	4.286 ± 5.121	12.632 ± 18.264	1.223 ± 1.832	5.464 ± 4.533
Missing data	13 (15.85)						
Work exposures							
Deny	46 (40.25)	p=0.501 3.001 ± 3.456	p=0.620 2.370 ± 1.858	p=0.020 2.159 ± 3.033	p=0.455 9.090 ± 10.143	p=0.302 0.727 ± 1.154	p=0.218 5.614 ± 8.037
Report	36 (43.90)	3.315 ± 4.440	4.893 ± 9.363	5.076 ± 7.727	10.696 ± 13.886	0.599 ± 0.542	7.251 ± 9.202

levels of miR-31-3p were higher in patients positive for work exposures but statistical significance was not reached ($p=0.081$). We found significant association between miR-31-5p ($p=0.048$) and miR-145-5p ($p=0.035$) and tumor stage. MiR-145-5p was significantly decreased with progression of T stage ($p=0.035$), positive nodal metastasis ($p=0.007$), and heavy tobacco smoking ($p=0.050$). MiR-222-3p was significantly associated with gender ($p=0.023$), positive family history ($p=0.015$), work exposures ($p=0.020$), and borderline associated with age ($p=0.052$). Decreased expression of miR-424-5p was associated with positive nodal metastasis ($p=0.012$), and a trend was shown for association with tumor localization ($p=0.015$), as higher expression was reached in subglottic tumors. lncRNA MALAT1 was positively associated with age ($p=0.024$), whereas lncRNA HOTAIR with positive family history ($p=0.031$). However, miR-21-3p, miR-155-5p, miR-196a-5p, miR-210-3p and miR-221-3p did not show any significant association with the explored clinical features.

Discussion

The aim of the current study was to explore and analyze the association between the expression levels of certain isoforms of commonly deregulated miRNAs and lncRNAs and the clinicopathological features of Bulgarian patients with laryngeal squamous cell carcinoma.

Development of LSCC involves various genetic and epigenetic changes, including aberrant expression of ncRNAs, including miRs and lncRNAs. ncRNAs have an important, but still widely uncovered regulatory role in adjusting the canonical cascades of laryngeal carcinogenesis. The obtained results showed statistically different expression of all studied ncRNAs in LSCC tumour tissue in comparison to adjacent normal samples, which suggests their involvement in laryngeal carcinogenesis. Still most promising miRs associated with patient features were: miR-21-5p, miR-31-3p, miR-31-5p, miR-145-5p, miR-222-3p, miR-424-5p, and lncRNA HOTAIR.

Intriguing for the first time, we found that the expression levels of miR-21-5p, miR-222-3p and lncRNA HOTAIR were statistically significantly associated with positive family cancer history of the LSCC patients. Family history is an important risk factor in cancer, however knowledge about ncRNA expression related to family history is still scarce. In our study, LSCC patients who were positive to family cancer history, reported other solid tumours, including head and neck cancers (thyroid, lip, throat), lung, breast, bladder, kidney, uterus, stomach, prostate, leukemia, duodenum and colo-

rectal cancer in their relatives. We suggest that the expression levels of miR-21-5p, miR-222-3p and lncRNA HOTAIR is potentially associated with aberrantly expressed target mRNA. One of the common targets of miR-21 and miR-222 is PTEN, which often is deregulated in various tumours, including head and neck neoplasms, due to somatic mutation, deletions or epigenetic silencing [15], and is associated with family cancer [16]. In addition, two lncRNA HOTAIR polymorphisms are published, rs7958904C>G and rs4759314G>A, that lead to higher expression associated with other family solid tumours [17]. Previously, miRs expression in patients with and without family breast cancer was explored, but statistically different expression was not found, including miR-21 [18], whereas in another study miR-155 was found significantly deregulated in patients with family lung cancer [19]. Our data could promote additional investigation on aberrant expression of promising ncRNAs in cases with family cancer history.

In addition to previous findings, miR-222-3p expression levels were marginally positively associated with aging, whereas lncRNA MALAT1 was significantly negatively associated with age. MiR-222 is reported as “aging miRNA” [20], whereas downregulation of lncRNA MALAT1 in relation to age was not published previously and deserves further investigation. The factor “age” should also be taken into account when comparing results for these ncRNAs between studies, in addition to ethnicity and clinical characteristics. MiR-222-3p, X-chromosome-related miR, has been previously reported to be associated with gender. Elevated expression was observed in male compared to female patients in a study of Khalifa et al in 2016. Significant relationship between FOXP3 promoter polymorphism rs3761548A/C, the expression of miR-222 and gender effect was detected [21]. lncRNA MALAT1 expression levels were not associated previously with gender, but still both markers should be validated additionally in larger female LSCC group.

MiR-424-5p was upregulated in LSCC patients with positive and negative node status, but the results showed significant negative association with positive nodal metastasis group. Our finding is in line with previously described decreased expression levels of miR-424-5p in nodal metastatic esophageal squamous cell carcinoma (ESCC) and epithelial ovarian cancer [22,23]. However, we identified for the first time strong association between the expression levels of miR-424-5p and laryngeal tumour location. We showed gradual increase in the expression level from supraglottis, through glottis and subglottis. Subglottic nodal metastases are not common in LSCC and they are less metastatic

tumours in comparison to supraglottic, which are more lymph-enriched [24]. We could assume that marked overexpression of miR-424-5p could be potential protective marker for nodal metastasis and invasion. The other isoform, miR-424-3p, is reported as one of the most positively correlated supraglottic miRs [25], whereas we investigated miR-424-5p in all LSCC locations, and found its expression was significantly negatively associated with supraglottic laryngeal location. We believe that aberrant miR-424 isoforms expression could potentially be associated with LSCC specific location, and could be used as surrogate localization marker in clinical practice.

In the current study, miR-31-3p and miR-145-5p levels were negatively associated with nodal metastasis. Moreover, miR-145-5p was downregulated with elevated T-stage, and heavy smoking, while the “active” isoform miR-31-5p was overexpressed in advanced T-stage tumors. Paclitaxel sensitivity and drug resistance are related to aberrantly expressed miR-31 in investigated LSCC patients [26], and colorectal cancer (CRC) patients [27], whereas downregulation of miR-145 is found to be related with development and progression of CRC and increases the sensitivity to cetuximab in CRC independently of KRAS status through downregulating BCL2 expression [28,29]. These two miRs could be suggested as potential markers of treatment and prognosis.

miR-21 and miR-31 are reported as aberrantly expressed miRs due to environment exposure, tobacco smoking and the development of cancer [30]. Our results confirm more specifically miR-21-5p isoform association with harmful work exposures, whereas the expression levels of miR-31-3p isoform were higher in patients positive for harmful work exposures, but could not reach statistical significance. In addition, the positively associated miR-222 in LSCC patients who report work exposures ($p=0.020$) is in line with a previous report by Bollati et al in 2010. They investigated the changes in the expression levels of miR-222 as candidate biomarker in peripheral blood leukocytes obtained from 63 postexposure workers [31].

Our data indicated elevated expression of miR-21-5p associated with LSCC poor tumour differentiation, whereas on the contrary, upregulated levels of “passenger” miR-31-3p were more typical for well and moderately differentiated LSCC, suggesting its strong oncogenic activity during early stages of carcinogenesis. Our findings are in agreement with previously published data about miR-31, but not miR-21 association with tumour differentiation in verrucous head and neck cancer [32]. Both, miR-21 and miR-31, are among the most

investigated miRs, widely expressed in most SCCs, including laryngeal cancer [33], but still the knowledge about their isoform association with different clinicopathological features in LSCC is elusive.

The rest of the investigated miR isoforms (miR-21-3p, miR-155-5p, miR-196a-5p, miR-210-3p and miR-221-3p) were aberrantly expressed in LSCC tumor tissue, but were not associated with any of the clinicopathological features.

Recent studies reported that miR-21-3p is an oncogene which is highly associated with microsatellite instability and promotes cellular mobility through epithelial-mesenchymal transition [34], and is shown to be significantly associated with the depth of tumor invasion, nodal metastasis and clinical stage in gastric cancer [35]. Previously, miR-155 was investigated in 63 laryngeal tumor specimens and the results showed significant positive correlation to T-stage and poor tumor differentiation, and it was suggested as oncogene promoting laryngeal invasiveness through targeting SOCS1 and STAT3 [36]. One possible explanation of the discrepant results could be that in this study the expression levels in tumors were compared to only 21 laryngeal normal tissue [36], while in the current study we compared equal numbers of 63 tumor and adjacent normal tissues. In our study samples, RQ levels of miR-196 were extremely elevated with no significant association with patients' clinical features. In 2013 in the study of Saito et al, miR-196a was presented as the most promising laryngeal cancer biomarker with high expression levels in early T-stages [35]. We also found higher miR-196a-5p levels in T1 and T2 stages, which could support its active role in very early stages of LSCC disease progression. In renal cancer, miR-210 is associated with cancer metastasis, whereas miR-221 is suggested as suitable biomarker for lower cancer-specific treatment [38], but we did not find any significant association with the investigated features. The obtained results could be due to limitations of the sample size.

In summary, based on the existing studies and the current observations, we could not clarify the role of miR-21-3p, miR-155-5p, miR-196a-5p, miR-210-3p and miR-221-3p in laryngeal carcinogenesis, which could be proven by further more extensive investigations. However, at this stage they could not be chosen as suitable markers for LSCC diagnosis and prognosis.

We found that MALAT1 was underexpressed in the investigated LSCC tumour tissues. In many articles MALAT1 was previously described as a cancer-promoting and metastasis-promoting lncRNA, while other recent reports suggested a tumor-suppressing role of MALAT1 [39-41].

The findings in the literature are controversial about the role of MALAT1 in cancer. In the study of Xu et al in 2015 in breast cancer the levels of MALAT1 were downregulated, associated with short relapse-free survival and suggested for target therapy biomarker [42]. In addition, downregulation of MALAT1 was predicted as stemness and proliferative marker associated with activation of LTBP3 gene and suppressed expression of Sox2 and Nestin in glioma mesenchymal stem SHG139S cell line. Moreover, decreased levels of MALAT1 led to activation of ERK/MAPK signaling pathway and promoted proliferation [43]. On the other hand, in multiple myeloma, knockdown of MALAT1 induced apoptosis, but not proliferation, through the activation of mitochondrial-controlled apoptosis by upregulation of Bax, Caspase-3/-9 expression and downregulation of Bcl-2 [44]. In 2016 Fang et al showed that knockdown of MALAT1 in tongue squamous cell carcinoma cell lines (CAL27 and SCC-25) is associated with upregulation of SPRR2A protein, which influenced distant metastasis of TSCC [45]. Controversially, previous investigations in 2015 of Pang et al found elevated RQ levels of MALAT1 in pancreatic cancer, and its oncogenic potential was discussed for targeted strategy [46].

There are only limited studies on MALAT1 expression in laryngeal cancer. Intriguingly Feng et al published in 2012 aberrantly upregulated expression of MALAT1 in 72 LSCC specimens, associated with poor tumour differentiation and active proliferation [47].

We have carefully examined the experimental settings and approaches that have been used to study MALAT1 and could speculate that the discrepancies of the observations between studies could be due to various reasons, some related to experimental and statistical issues. In the study of Feng et al 18S was used as endogenous control [47], while in our study we compared the levels of MALAT1 to ACTB control, which was suggested recently as suitable endogenous control [48]. Zhou et al published in 2015 upregulation of MALAT1 expression in 54 randomly collected OSCC samples with only 12 normal mucosa samples, but they used GAPDH endogenous control. The tumour and normal samples were not from the same patients and the numbers of normal and tumour samples were not equal [49]. The statistical power of such sample to detect true differences in expression is very limited.

In addition, several molecular mechanisms by which MALAT1 regulates tumor progression and metastasis have been suggested, including its action as competitive endogenous (ceRNA) or decoy

miRs; its interaction with the Polycomb repressive complex 2 (PRC2), catalyzing histone methylation and playing important role in transcriptional repression and cancer; binding and inactivating TEAD transcription factor; regulating multiple signaling pathways in cancer such as Hippo-YAP, PI3K-AKT, MAPK, WNT, and NF- κ B pathways [39].

It is still unclear if the conserved and highly abundant lncRNA in normal tissues MALAT1 acts as tumour suppressor or tumour inducer. Comparing the obtained and published results, we could conclude that the expression of MALAT1 may differ in different tissues, cancer types and genetic backgrounds. Unraveling its precise mechanism of action and contribution to laryngeal cancer needs more comprehensive studies.

In conclusion, we explored the expression levels of 11 miRs and two lncRNAs (MALAT1 and HOTAIR) in LSCC and adjacent normal tissue in Bulgarian patients, revealing new associations between the studied ncRNAs and clinicopathological features. We demonstrated new potential association between family history and the expression levels of miR-21-5p, miR-222-3p and lncRNA HOTAIR, downregulation of MALAT1 with age and potential gender effect, upregulation of “passenger” miR-31-3p in well and moderately differentiated LSCC tumours, and potential role of miR-424-5p isoform in LSCC location screening. Our research confirms some previous findings and shows controversial results to other investigations. MiR-145-5p was negatively related to T-stage and tobacco smoking, while miR-31-3p association with nodal metastasis could serve as potential marker for cancer treatment. We validated the potential role of miR-21-5p and miR-222-3p with harmful work exposures.

These results contribute to better understand the mechanisms of carcinogenesis in laryngeal cancer as well as to elucidate the potential role of ncRNA as biomarkers in the clinical practice.

Acknowledgement

This study was funded by Medical University – Sofia projects: no. 312/15.01.2015 (Contract no. 15-D); no. 552/21.01.2016 (contract no. 12-D); no. 8553/12.12.2016 (Contract no. D-137), and supported by infrastructure grant DUNK01-2/2009 and no. H13/29 (Contract DH13/12/20.12.2017) by National Science Fund, Ministry of Education and Science of Bulgaria. Special thanks to all patients enrolled in this study.

Conflict of interests

The authors declare no conflict of interests.

References

1. Ferlay J, Soerjomataram I, Dikshit R et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer* 2015;136:E359-86.
2. Valerianova Z, Atanasov T, Vukov M. Cancer incidence in Bulgaria, 2014 & 2015, Vol XXV, 2017.
3. Ferlay J, Colombet M, Soerjomataram I et al. Estimating the global cancer incidence and mortality in 2018: GLOBOCAN sources and methods. *Int J Cancer* 2019;144:1941-53.
4. Steuer CE, El-Deiry M, Parks JR, Higgins KA, Saba NF. An update on larynx cancer. *CA Cancer J Clin* 2017; 67:31-50.
5. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2016. *CA Cancer J Clin* 2016; 66:7-30.
6. Okazaki Y, Furuno M, Kasukawa T et al. Analysis of the mouse transcriptome based on functional annotation of 60,770 full-length cDNAs. *Nature* 2002;420:563-73.
7. Lagos-Quintana M, Rauhut R, Lendeckel W, Tuschl T. Identification of novel genes coding for small expressed RNAs. *Science* 2001;294:853-8.
8. Pradhan AK, Emdad L, Das SK, Sarkar D, Fisher PB. The Enigma of miRNA Regulation in Cancer. *Adv Cancer Res* 2017;135:25-52.
9. Ha M, Kim VN. Regulation of microRNA biogenesis. *Nat Rev Mol Cell Biol* 2014;15:509-24.
10. Choudhry H, Harris AL, McIntyre A. The tumour hypoxia induced non-coding transcriptome. *Mol Aspects Med* 2016;47-48:35-53.
11. Medina PP, Nolde M, Slack FJ. OncomiR addiction in an in vivo model of microRNA-21-induced pre-B-cell lymphoma. *Nature* 2010;467:86-90.
12. Kozomara A, Birgaoanu M, Griffiths-Jones S. miRBase: from microRNA sequences to function. *Nucleic Acids Res* 2019;47:155-62.
13. Fang S, Zhang L, Guo J et al. NONCODEV5: a comprehensive annotation database for long non-coding RNAs. *Nucleic Acids Res* 2018;46 (Database issue):308-14.
14. Livak KJ, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C(T)) Method. *Methods* 2001; 25:402-8.
15. Snietura M, Jaworska M, Mlynarczyk-Liszka J et al. PTEN as a Prognostic and Predictive Marker in Postoperative Radiotherapy for Squamous Cell Cancer of the Head and Neck. *PLoS One* 2012;7:e33396.
16. Bakarakos P, Theohari I, Nomikos A et al. Immunohistochemical study of PTEN and phosphorylated mTOR proteins in familial and sporadic invasive breast carcinomas. *Histopathology* 2010;56:876-82.
17. Wu H, Shang X, Shi Y et al. Genetic variants of lncRNA HOTAIR and risk of epithelial ovarian cancer among Chinese women. *Oncotarget* 2016;7:41047-52.
18. Han JG, Jiang YD, Zhang CH et al. A novel panel of serum miR-21/miR-155/miR-365 as a potential diagnostic biomarker for breast cancer. *Ann Surg Treat Res* 2017;92:55-66.
19. Fawzia Khalil Ibrahim, Randa Ali-Labib, Iman Hassan Galal, Hala Moustafa Mahmoud. MicroRNA-155 expression in exhaled breath condensate of patients with lung cancer. *Egypt J Chest Dis Tuberculosis* 2017; 66:687-91.
20. Zhang H, Yang H, Zhang C et al. Investigation of MicroRNA Expression in Human Serum During the Aging Process. *J Gerontol A Biol Sci Med Sci* 2015; 70:102-9.
21. Khalifa O, Pers YM, Ferreira R et al. X-Linked miRNAs Associated with Gender Differences in Rheumatoid Arthritis. *Int J Mol Sci* 2016;17:pii: E1852
22. Wang F, Wang J, Yang X, Chen D, Wang L. MiR-424-5p participates in esophageal squamous cell carcinoma invasion and metastasis via SMAD7 pathway mediated EMT. *Diagn Pathol* 2016;11:88.
23. Liu J, Gu Z, Tang Y, Hao J, Zhang C, Yang X. Tumour-suppressive microRNA-424-5p directly targets CCNE1 as potential prognostic markers in epithelial ovarian cancer. *Cell Cycle* 2018;17:309-18.
24. Ma H, Lian M, Feng L et al. Factors contributing to lymph node occult metastasis in supraglottic laryngeal carcinoma cT2-T4 N0M0 and metastasis predictive equation. *Chin J Cancer Res* 2014;26:685-91.
25. Sun X, Song Y, Tai X, Liu B, Ji W. MicroRNA expression and its detection in human supraglottic laryngeal squamous cell carcinoma. *Biomed Rep* 2013; 1:743-6.
26. Li P, Liu H, Wang Z et al. MicroRNAs in laryngeal cancer: implications for diagnosis, prognosis and therapy. *Am J Transl Res* 2016;8:1935-44.
27. Sur D, Cainap C, Burz C et al. The role of miRNA -31-3p and miR-31-5p in the anti-EGFR treatment efficacy of wild-type K-RAS metastatic colorectal cancer. Is it really the next best thing in miRNAs? *JBUON* 2019;24:1739-46.
28. Liu Q, Yang W, Luo Y, Hu S, Zhu L. Correlation between miR-21 and miR-145 and the incidence and prognosis of colorectal cancer. *JBUON* 2018;23:29-35.
29. Zhang Y, Wang J. MicroRNAs are important regulators of drug resistance in colorectal cancer. *Biol Chem* 2017;398:929-38.
30. Vrijens K, Bollati V, Nawrot TS. MicroRNAs as potential signatures of environmental exposure or effect: a systematic review. *Environ Health Perspect* 2015;123:399-411.
31. Bollati V, Marinelli B, Apostoli P et al. Exposure to Metal-Rich Particulate Matter Modifies the Expression of Candidate MicroRNAs in Peripheral Blood Leukocytes. *Environ Health Perspect* 2010;118:763-8.
32. Odar K, Boštjančič E, Gale N, Glavač D, Zidar N. Differential expression of microRNAs miR-21, miR-31, miR-203, miR-125a-5p and miR-125b and proteins PTEN and p63 in verrucous carcinoma of the head and neck. *Histopathology* 2012;61:257-65.
33. Wang P, Fu T, Wang X, Zhu W. Primary, study of miRNA expression patterns in laryngeal carcinoma by microarray. *Lin Chung Er Bi Yan Hou Tou Jing Wai Ke Za Zhi* 2010;24:535-8.
34. Jiao W, Leng X, Zhou Q et al. Different miR-21-3p isoforms and their different features in colorectal cancer. *Int J Cancer* 2017;141:2103-11.
35. Wen Xu, Muqing Yang, Ming Gao, Wei Wu, Zhewei Fei.

- MiR-21-3p and miR-21-5p in tumor tissue as diagnostic biomarkers for gastric cancer. *Int J Clin Exp Pathol* 2016;9:7195-201.
36. Zhao XD, Zhang W, Liang HJ, Ji WY. Overexpression of miR -155 Promotes Proliferation and Invasion of Human Laryngeal Squamous Cell Carcinoma via Targeting SOCS1 and STAT3. *PLoS One* 2013;8:e56395.
 37. Saito K, Inagaki K, Kamimoto T et al. MicroRNA-196a Is a Putative Diagnostic Biomarker and Therapeutic Target for Laryngeal Cancer. *PLoS One* 2013; 8:e71480.
 38. Dias F, Teixeira AL, Ferreira M et al. Plasmatic miR-210, miR-221 and miR-1233 profile: potential liquid biopsies candidates for renal cell carcinoma. *Oncotarget* 2017;8:103315-26.
 39. Sun Y, Li MA. New Insights into Long Non-Coding RNA MALAT1 in Cancer and Metastasis. *Cancers (Basel)* 2019;11:216.
 40. Zhang Y, Wang T, Huang HQ, Li W, Cheng XL, Yang J. Human MALAT-1 long non-coding RNA is overexpressed in cervical cancer metastasis and promotes cell proliferation, invasion and migration. *JBUON* 2015;20:1497-503.
 41. Wanga Y, Wua C, Zhang C et al. TGF- β -induced STAT3 overexpression promotes human head and neck squamous cell carcinoma invasion and metastasis through malat1/miR-30a interactions. *Cancer Lett* 2018;436:52-62.
 42. Xu S, Sui S, Zhang J et al. Downregulation of long noncoding RNA MALAT1 induces epithelial-to-mesenchymal transition via the PI3K-AKT pathway in breast cancer. *Int J Clin Exp Pathol* 2015; 8:4881-91.
 43. Han Y, Zhou L, Wu T et al. Downregulation of lncRNA-MALAT1 Affects Proliferation and the Expression of Stemness Markers in Glioma Stem Cell Line SHG139S. *Cell Mol Neurobiol* 2016;36:1097-107.
 44. Liu H, Wang H, Wu B et al. Down-regulation of long non-coding RNA MALAT1 by RNA interference inhibits proliferation and induces apoptosis in multiple myeloma. *Clin Exp Pharmacol Physiol* 2017;44:1032-41.
 45. Fang Z, Zhang S, Wang Y et al. Long non-coding RNA MALAT-1 modulates metastatic potential of tongue squamous cell carcinomas partially through the regulation of small proline rich proteins. *BMC Cancer* 2016;16:706.
 46. Pang EJ, Yang R, Fu XB, Liu YF. Overexpression of long non-coding RNA MALAT1 is correlated with clinical progression and unfavorable prognosis in pancreatic cancer. *Tumour Biol* 2015;36:2403-7.
 47. Feng J, Tian L, Sun Y et al. Expression of long non-coding ribonucleic acid metastasis-associated lung adenocarcinoma transcript-1 is correlated with progress and apoptosis of laryngeal squamous cell carcinoma. *Head Neck Oncol* 2012;4:46.
 48. Chang J, Xu W, Du X, Hou J. MALAT1 silencing suppresses prostate cancer progression by upregulating miR-1 and downregulating KRAS. *Onco Targets Ther* 2018;11:3461-73.
 49. Zhou X, Liu S, Cai G et al. Long Non Coding RNA MALAT1 Promotes Tumor Growth and Metastasis by inducing Epithelial-Mesenchymal Transition in Oral Squamous Cell Carcinoma. *Sci Rep* 2015;5:15972.